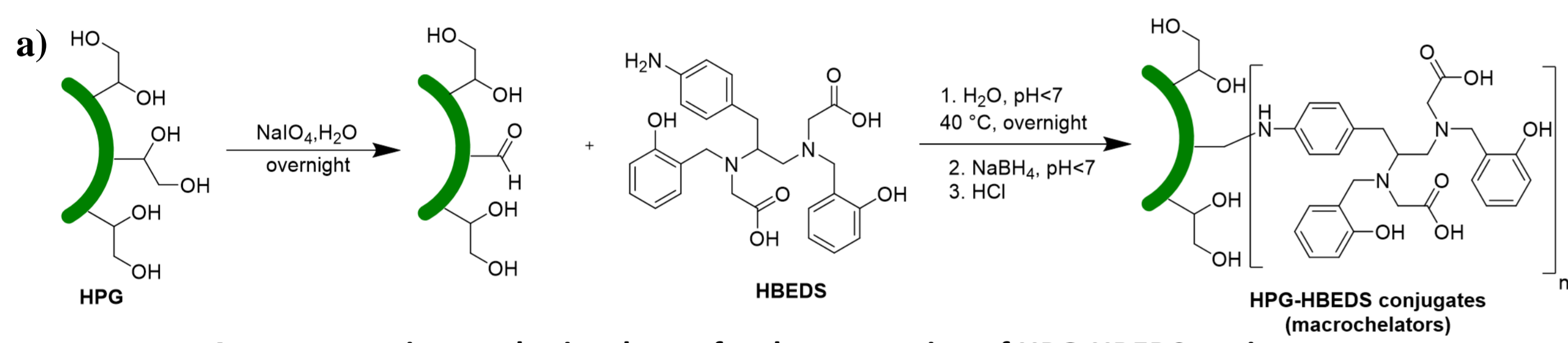


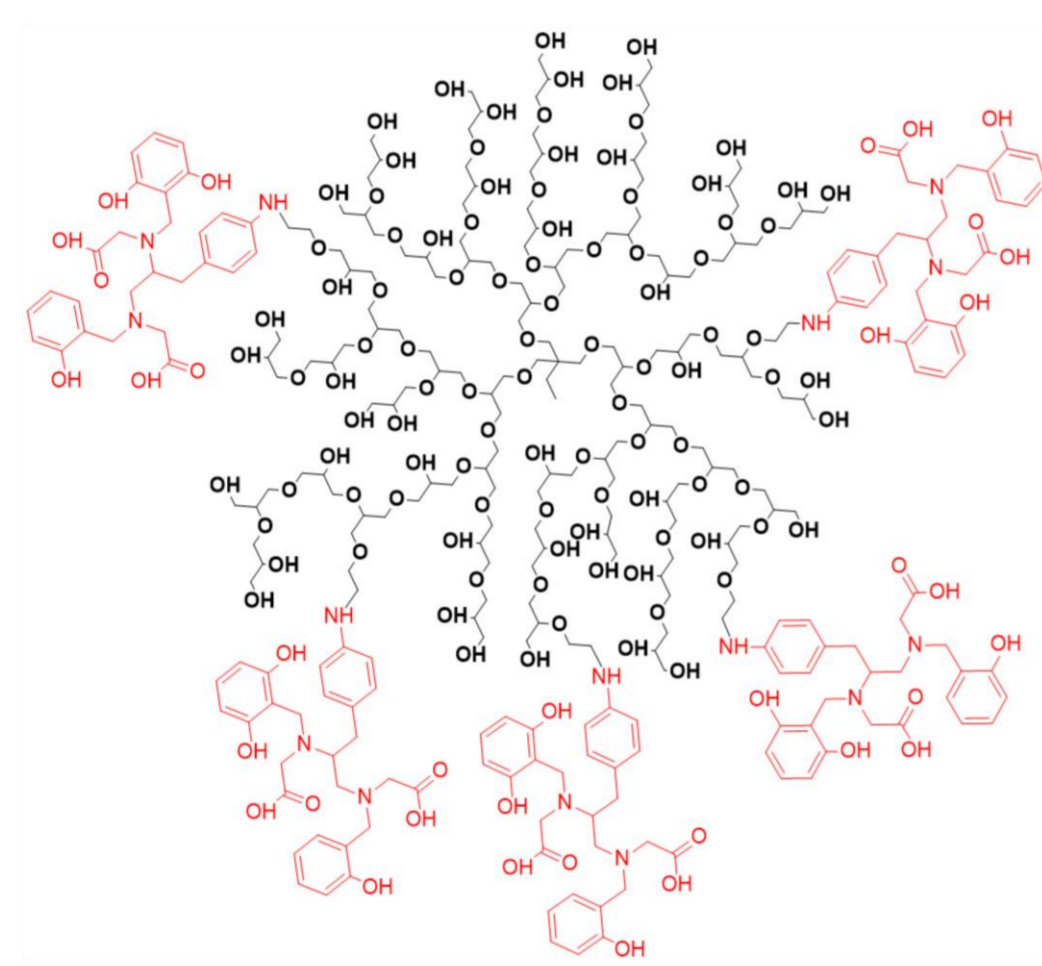
Introduction

- Millions of resistant nosocomial infections occur every year
- Severe lack of new antibiotics being developed + biofilms confer even more resistance
- These infections become systemic & require harsher antibiotics → worse side-effects
- Unique approaches needed to combat this persistent problem.
- Our method takes advantage of iron & its role in infection.**
 - Upon infection → war for iron acquisition between the host & bacteria
 - Bacteria secrete small iron binding molecules, *siderophores*, to bind + uptake iron
 - Siderophores vary in structure, size, and $K_{complex}$
 - Iron → essential nutrient for most bacterial metabolisms
 - Iron sequestration should limit/eradicate bacteria growth**
- Iron sequestration has been attempted with various siderophores
 - Literature results poor
 - Easy uptake & high dosage for tested small chelators
 - Macromolecular approaches limit uptake
 - complex synthesis + solubility issues
- Ideal macrochelator requirements:
 - Be able to compete with siderophores ($K_{complex}$ between 10^{30} to 10^{40})
 - Not be up-taken by bacteria
 - Won't harm host
 - Highly soluble
 - Easily Synthesized
- Our Approach:**
 - High-molecular weight polymeric scaffold (Hyper-Branched Polyglycerol, *HPG*) conjugated to a modified high-affinity iron chelator, *HBEDS*
 - Investigated inhibition/eradication of biofilms + effect of molecular weight of macrochelator

Approach



- A representative synthetic scheme for the generation of HPG-HBEDS conjugates



- Structure HPG-HBEDS conjugates. Black is the HPG scaffold. Red is the conjugated HBEDS chelator

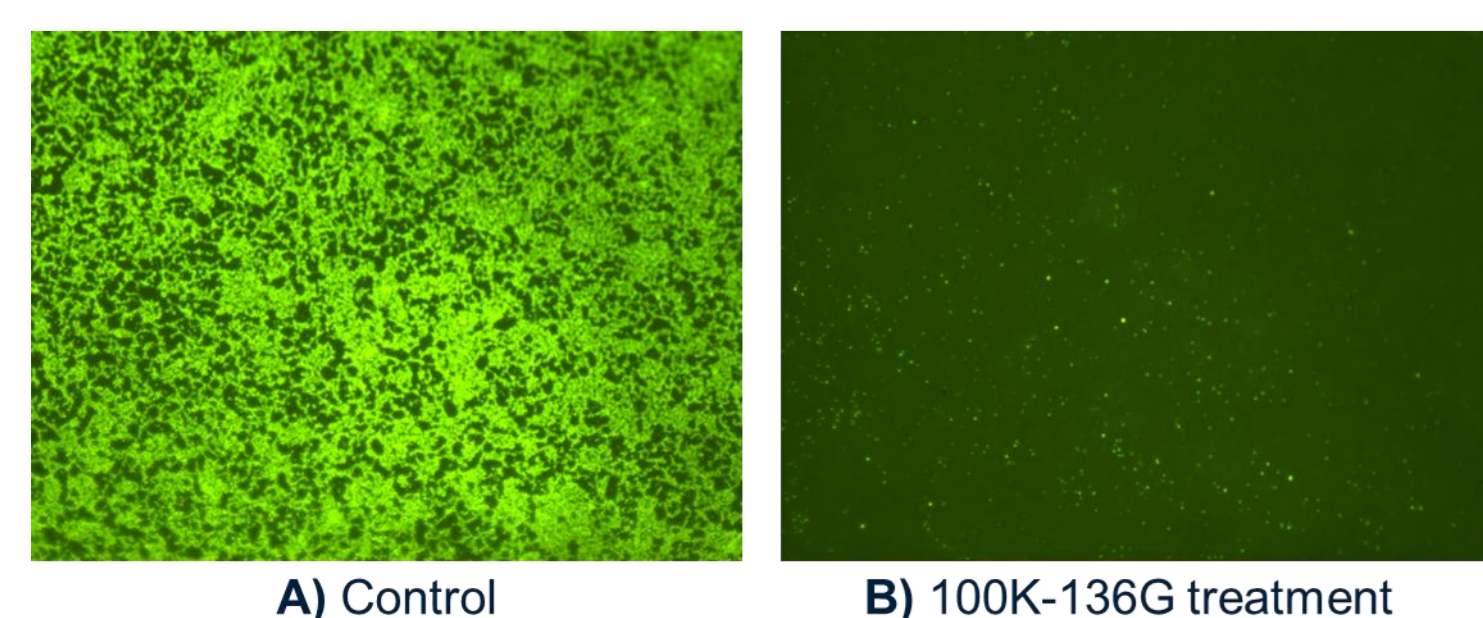
Methods

- Library of macrochelators created by conjugating hyperbranched polyglycerol (HPG) (100kDa and 31kDa) to a modified small molecule chelator, *HBEDS*, via reductive amination
- Macrochelators characterized via NMR, IR, & UV-Vis spectroscopy to confirm conjugation & number of HBEDS groups
- S. aureus* lux strain (Xen36 Lux) was main strain studied
 - Biofilm eradication assays performed by growing a biofilm first (100ul of 10^6 CFU culture) for 24 hours in polypropylene 96 well plate in 50% TSB media
 - Media was drained, water washed 3x, & treated with macrochelator
 - Crystal violet assay conducted as described in literature
 - For inhibition assays, conjugate was added with bacteria on day one
- Modified eradication protocol used for confocal studies with syto9 stain for imaging
 - Our molecules labelled with AZDye 405 cadaverine via reductive amination.
- Data normalized to control
- Error bars represent standard deviations of 3 biological replicates, N=3

Results

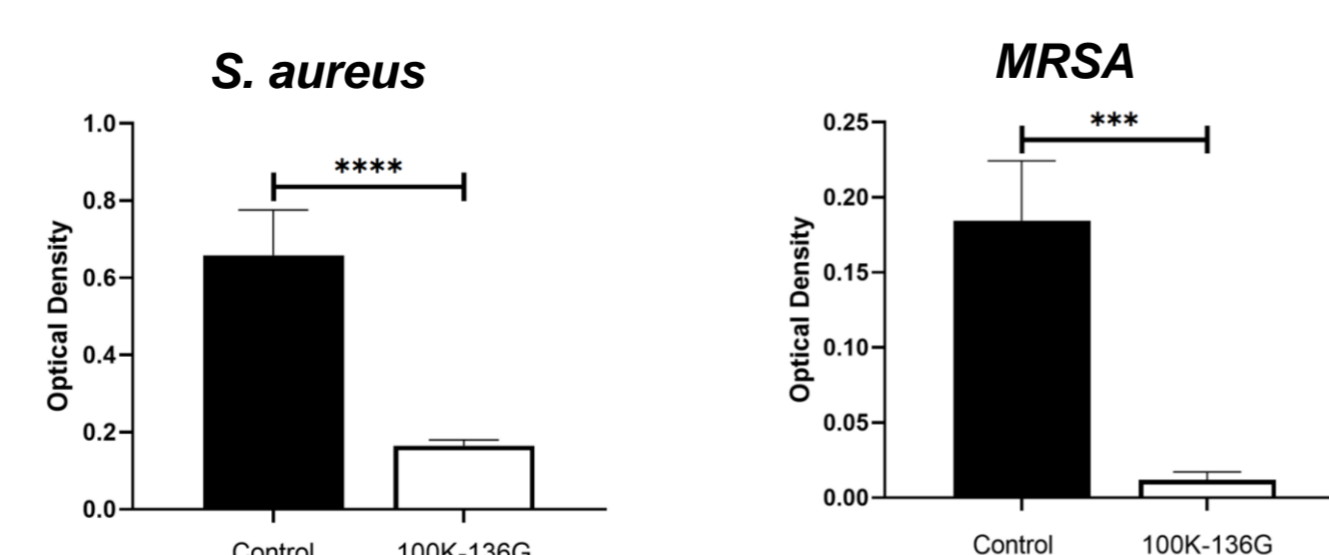
Destruction of Pre-formed Biofilms

1. Fluorescence Microscopy of *S. aureus* biofilm after conjugate treatment



- Image shows reduction in biofilm after conjugate treatment relative to untreated control

2. Crystal violet assay shows biofilm eradication

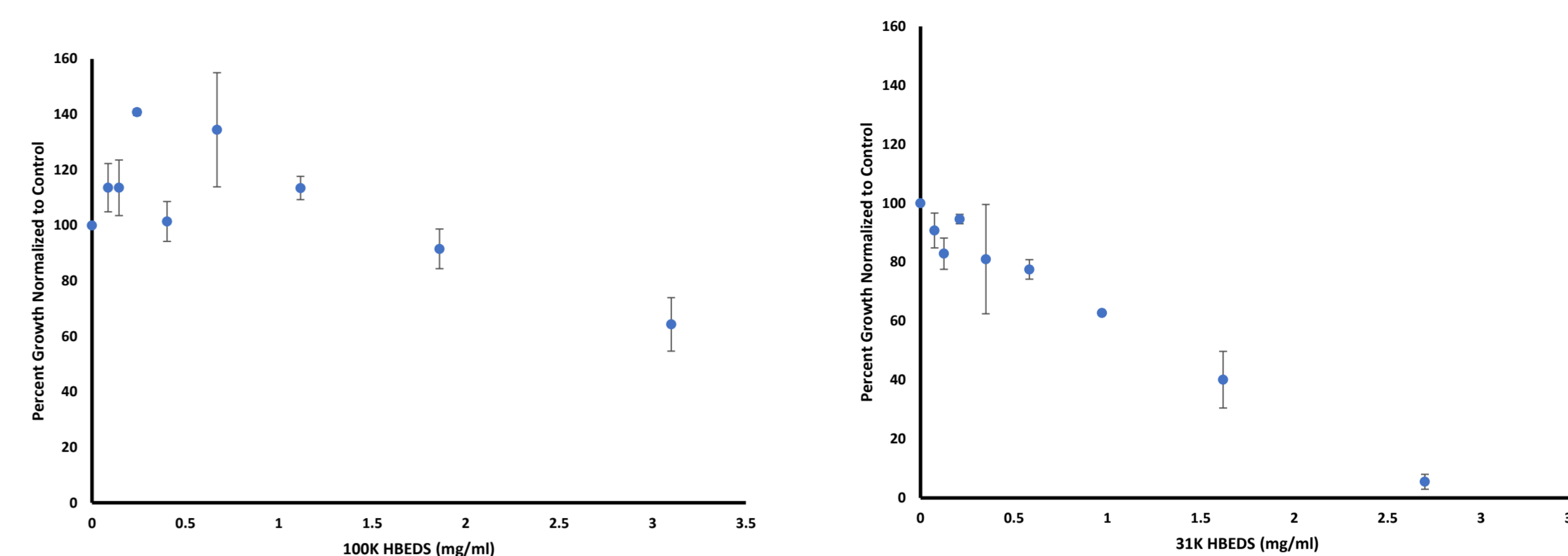


- Conjugate exhibited biofilm killing against both *S. aureus* and MRSA of up to 75% & 94%, respectively

Results

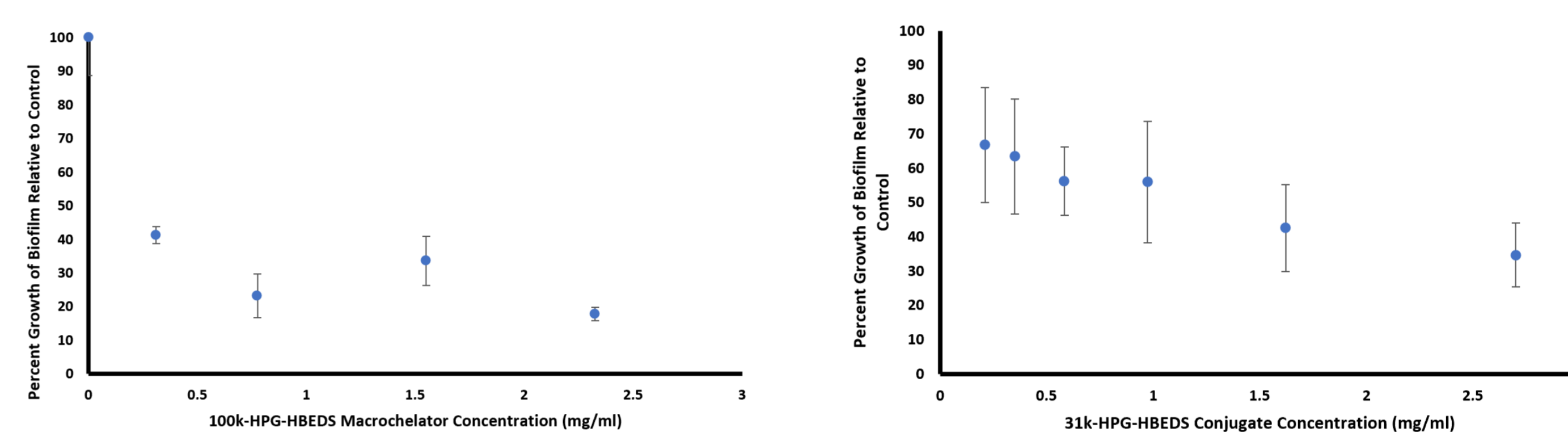
Effect of Molecular Weight of Chelators on Inhibition and Eradication of *S. aureus* Biofilm

3. Biofilm inhibition of *S. aureus* in presence of macrochelators of 2 different molecular weights (100kDa and 31kDa respectively) at different concentrations



- 31kDa variant seems to perform much better at biofilm inhibition than the 100kDa variant.

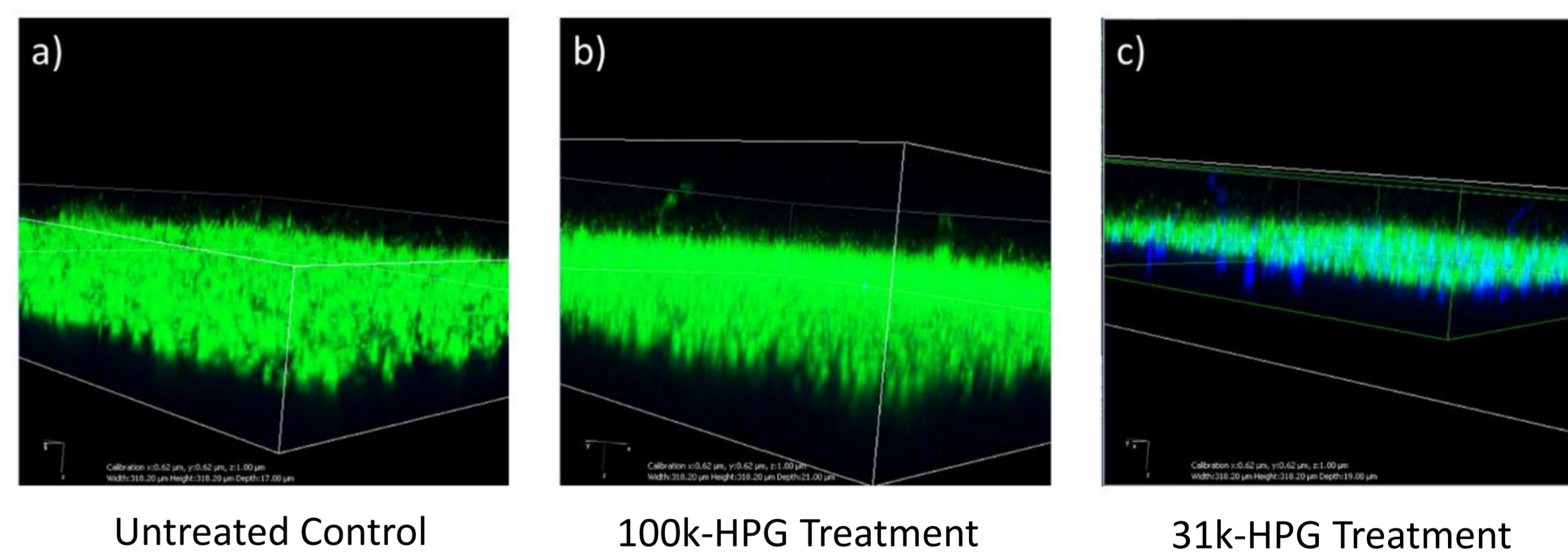
4. Biofilm eradication of *S. aureus* in presence of macrochelators of 2 different molecular weights (100kDa and 31kDa respectively) at different concentrations



- 100kDa variant is more effective than the 31kDa variant at lower concentrations.

Effect of Scaffold Molecular Weight on Diffusion of Macrochelator into Biofilm

5. Confocal study of polymer scaffold diffusion into biofilm after 16 hours



- Clearly shown that 31kDa scaffold penetrates biofilm while 100kDa scaffold does not

Conclusions

- Our macrochelators have biofilm eradication properties against MRSA and *S. aureus*.
- 31kDa conjugate was much less efficient
- 100 kDa backbone did not penetrate the biofilm
- 31 kDa backbone penetrated the biofilm but also inserted itself underneath
 - Biofilm penetration may account for differences in respective eradication abilities
- Studies show importance of further understanding the microenvironment of biofilms in order to effectively combat them
- More studies underway on exploring these macrochelators further in the context of biofilm eradication both as solo therapeutics or as adjuvants.

References and Acknowledgements

- Abbina, S.; Gill, A.; Mathew, S.; Abbasi, U.; and Kizhakkedathu, J.N.; *ACS Applied Materials & Interfaces* 2020 12 (34), 37834-37844